



Transfection reagent

HYPE-CHO™

Transfection Kit

Achieve High Yield Protein Expression in CHO cells

Protocol

HYPE-CHO Quick Protocol for CHO, CHO...

To find the ideal conditions, HYPE-CHO reagent must be tested at ratios **1 $\mu\text{L}/\mu\text{g}$, 2 $\mu\text{L}/\mu\text{g}$ and 3 $\mu\text{L}/\mu\text{g}$** (μL of Hype-CHO / μg of DNA). For the DNA quantity, we suggest to test from **1.5 μg to 2 μg** of DNA per mL of culture medium. The BCHO reagent must be used at **1/100** of the total volume.

1h before transfection, dilute your cells at 1×10^6 cells per mL*

| | | |
|----------|----------------------|-----------------------|
| 1 | 125 mL Bottle | 1 L Bottle |
| | 30 mL culture medium | 250 mL culture medium |
| | 30 x 10^6 cells | 250 x 10^6 cells |

Prepare 3 identical tubes of DNA



| | | |
|----------|--|--|
| 2 | 125 mL Bottle | 1 L Bottle |
| | 45 or 60 μg in 500 μL of serum-free medium or buffer* x3 | 375 or 500 μg in 5mL of serum-free medium or buffer* x3 |

Prepare 3 tubes of HYPE-CHO (with 3 different amounts of reagent)



| | | | | |
|----------|----------------------|--|-----------------------|--|
| 3 | 125 mL Bottle | 1 L Bottle | | |
| | 45 μg DNA | 45 μL / 90 μL / 135 μL in 500 μL of serum-free medium or buffer* | 375 μg DNA | 375 μL /750 μL /1125 μL in 5mL of serum-free medium or buffer* |
| | 60 μg DNA | 60 μL /120 μL /180 μL in 500 μL of serum-free medium or buffer* | 500 μg DNA | 500 μL /1000 μL /1500 μL in 5mL of serum-free medium or buffer* |

Mix each DNA tube to each tube of HYPE-CHO and incubate 20min at RT*



Distribute each mix into the bottle containing cells, incubate under orbital shaking for 4h*



After 4 hours, add BCHO (1/100, 1X final) reagent directly into the cells*



30 mL Culture Medium - 320 μL of BCHO

250 mL Culture Medium - 2.7mL of B293

Incubate cells for 24h to 7 days under orbital shaking (or more if needed) at 37°C until evaluation of protein production

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Choose the optimal conditions (DNA quantity, DNA/HYPE-CHO ratio, BCHO)

* Please refer to the following section "Important Notes"

IMPORTANT NOTES – Before you begin

- ✓ HYPE-CHO™ Kit has been used and validated with cells from different origins (CHO, CHO-S, rCHO or any CHO-related cells in suspension). It is suitable for any kind of mammalian cells used to produce proteins. This Kit has been tested with several chemically defined media. It is compatible with any specific media for protein production. Do not use culture medium containing high antibiotic level (up to 0.5 X penicillin/streptomycin final concentration) or high Pluronic® surfactant concentration (up to 0.01% w/v final concentration) to avoid dramatic impact on protein production level.
- ✓ The instructions given represent protocols that were applied successfully with a variety of CHO cells growing in suspension and cultivated in chemically defined medium. Optimal conditions may vary depending on the nucleic acid, cell types, growth condition (medium, size of cell culture...). Therefore, we suggest optimizing the various parameters as described in the complete instruction manual. However, to obtain good data rapidly, you can start by following our rapid protocol as guidelines.
- ✓ **The use of BCHO** reagent is highly recommended yet optional. We observed, when using BCHO reagent, a large increase in protein expression with our CHO suspension cell model.
- ✓ 18-24 h before transfection, seed the cells to 0.6-0.8 x 10⁶ cells/mL and incubate on orbital shaker (~125 rpm) at 37°C, 8% CO₂. The day of transfection, dilute the cells to 1 x 10⁶ cells/mL (cell density should be about 0.8-1.2 x 10⁶ cells/mL).
- ✓ Allow reagents to reach RT and gently vortex prior to use.
- ✓ **Medium or buffer without serum & supplement** must be used for the preparation of complexes (DNA/Hype-CHO). Culture media such as MEM, DMEM or OptiMEM or buffers such as HBS or PBS are recommended. In contrast, we do not recommend RPMI for preparing the complexes.
- ✓ **Formation of complexes.** Add the DNA solution into the HYPE-CHO solution, mix gently by carefully pipetting up and down 3 to 5 times. Incubate the mixture for 20 minutes at room temperature. Do not vortex or centrifuge!
- ✓ Bioreactor, spinner, flasks or Erlenmeyer etc. can be used.

For additional information and protocols (optimization, scaling, co-transfection...) tips, troubleshooting or other applications



www.ozbiosciences.com

Any questions?



tech@ozbiosciences.com

HYPE-CHO Reagent | Specifications

| | |
|---------------------|--|
| Package content | HYC01500: 1.5 mL of HYPE-CHO + 5 mL of BCHO HYC03000: 2 x 1.5 mL of HYPE-CHO + 2 x 5 mL of BCHO HYC15000: 15 mL of HYPE-CHO + 50 mL of BCHO HYC30000: 2 x 15 mL of HYPE-CHO + 2 x 50 mL of BCHO |
| Shipping conditions | Room Temperature |
| Storage conditions | Store the HYPE-CHO transfection reagent and BCHO at 4°C upon reception |
| Shelf life | 1 year from the date of purchase when properly stored and handled |
| Product description | HYPE-CHO is a high efficiency transfection reagent specifically developed to achieve High Yield Protein Expression in CHO cells growing in suspension. |
| Important notice | For research use only. Not for use in diagnostic procedures |

1. Cells preparation

Cell culture maintenance: sub-culture the cells at a density of $0.5-2 \times 10^6$ cells/mL for each passage (48-72h). Avoid high cell density and keep cell growth conditions consistent for reproducibility. Cells must be actively dividing and in exponential growth, mainly as single cells before transfection for the maximum of efficiency.

18-24 h before transfection, dilute the cells to $0.6-0.8 \times 10^6$ cells/mL and incubate on orbital shaker (~125 rpm) at 37°C, 8% CO₂. The day of transfection, dilute the cells to 1×10^6 cells/mL* (cell density should be about $0.8-1.2 \times 10^6$ cells/mL). Transfer the volume of cells needed as described in Table 1.

* **NOTE:** for 1mL culture format, we recommend using a cell density of 2.5×10^5 cell/mL.

| Cell culture | | | DNA | | HYPE-CHO reagent | | BCHO (optional) | |
|------------------------------|---------------|---------------------|---------------------------|-----------------|------------------|-----------------|-----------------------------|-----------------|
| 10 ⁶ cells per mL | | | 1.5 µg/mL of cell culture | | 2 µL per µg DNA | | 1X final dilution 4 h later | |
| Culture volume | Culture Flask | Total cell Number* | Quantity µg | Dilution volume | Volume µL | Dilution volume | Volume µL | Dilution volume |
| 1 mL | NA | 1x10 ⁶ | 1.5 µg | 50 µL | 3 µL | 50 µL | 12 µL | 100 µL |
| 30 mL | 125 mL | 30x10 ⁶ | 45 µg | 0.6 mL | 90 µL | 0.6 mL | 320 µL | 1 mL |
| 250 mL | 1 L | 250x10 ⁶ | 375 µg | 5 mL | 750 µL | 5 mL | 2.7 mL | 10 mL |
| 1 L | 3 L | 1x10 ⁷ | 1.5 mg | 20 mL | 3 mL | 20 mL | 10.8 mL | 40 mL |

* The day of transfection cell density should be at 1×10^6 cells/mL.

Table 1: Suggested volumes of HYPE-CHO, BCHO and DNA quantity

2. DNA/HYPE-CHO complexes preparation

- HYPE-CHO*: Vortex the reagent and dilute the indicated quantity of HYPE-CHO (refer to Table 1) in 50 µL to 20 mL of culture medium without serum and supplement.
- DNA*: Dilute the indicated quantity of DNA (see Table 1) in 50 µL to 20 mL of culture medium without serum and supplement.
- Add the DNA solution to the HYPE-CHO solution and mix by carefully pipetting up and down. Incubate the mix at room temperature for 20 minutes. Do not vortex or centrifuge.

3. Transfection

- Add the HYPE-CHO / DNA complexes dropwise into cell culture bottle while gently swirling the flask to ensure a uniform distribution. Incubate the cell on orbital shaker (~125 rpm) at 37°C, 8% CO₂.
- Four hours later add BCHO – 1X final, directly to the vessel containing cells (refer to Table 1)
- Cultivate the cells under standard conditions for 1 to 7 days depending on the type of protein expression. No medium change is required during the incubation period.

IMPORTANT CONSIDERATIONS

The use of BCHO reagent is highly recommended yet optional. We observed, when using BCHO reagent, a large increase in protein expression with our CHO suspension cell model.

HYPE-CHO allows easy scaling up and scaling down - it is compatible with various size volumes and culture vessels. Simply adjust each reagent proportion to the volume of culture medium. The Table 1 shows recommended amounts of HYPE-CHO, DNA and BCHO for 1mL to 1L of cell culture medium. Since transfection efficiency is depending on the cell model (clone, growth conditions...) and the culture vessels (shaker, spinner flask, bioreactor...), we recommend performing an optimization procedure (refer to next section) before scaling up or down.

Optimization Protocol

Although high protein production can be achieved in CHO cells growing in suspension following the previous protocol, some optimizations may be required in order to obtain the maximum efficiency. For best results, we recommend to optimize two parameters:

- Quantity of HYPE-CHO reagent and DNA
- Cell culture conditions

1. HYPE-CHO reagent and DNA parameters optimization

HYPE-CHO reagent must be used in slight excess compare to DNA but the optimal ratio will depend on the cell model and culture conditions.

First step: Maintain a fixed quantity of DNA to 1.5 µg/mL of cell culture and then vary the amount of HYPE-CHO reagent from 1.5 to 3µL per µg of DNA (see step one - Table 2 first for example).

Second step: Once the ratio of HYPE-CHO to DNA has been optimized, keep it constant and vary the DNA quantity from 1 to 3 µg per mL of cell culture (see step two - Table 2 step for example).

| Step | Cell culture | | DNA | | HYPE-CHO reagent | | BCHO |
|----------|----------------|-----------------------|---------------|-----------------|-----------------------|-----------------|----------|
| | Culture volume | Total cell Number* | Quantity µg | Dilution volume | Volume µL | Dilution volume | Quantity |
| Step one | 30 mL | 30 x 10 ⁶ | 45 | 0.6 mL | 67,90,135 | 0.6 mL | 320 µL |
| | 250 mL | 250 x 10 ⁶ | 375 | 5 mL | 562, 750, 1125 | 5 mL | 2.7 mL |
| Step two | 30 mL | 30 x 10 ⁶ | 30, 60, 90 | 0.6 mL | Ratio from first step | 0.6 mL | 320 µL |
| | 250 mL | 2.5 x 10 ⁸ | 250, 500, 750 | 5 mL | Ratio from first step | 5 mL | 2.7 mL |

* The day of transfection cell density should be at 1 x 10⁶ cells/mL.

Table 2: Example for HYPE-CHO and DNA optimization

To test whether or not BCHO increases your protein production, we advise to use the previous optimized HYPE-CHO/DNA parameters in two conditions: one with and one without BCHO.

2. Cell culture conditions optimization

Efficient protein production is also highly dependent on the cell model. For instance, several parameters are critical to obtain the maximum efficiency such as cell suspension growth adaptation, culture medium and cell density (before and during transfection).

We recommend optimizing cell density. After setting up the best ratio of HYPE-CHO/DNA and the DNA quantity, test various cell densities from 0.5 to 2×10^6 cells/mL at the time of transfection - cells must be in their growth phase. The cells must be grown as single cells because extensive clumping at the time of transfection can reduce the quantity of protein produced. If necessary, vigorous vortexing for 10-30 seconds could be done for single cell growth recovering.

Additional products for bioproduction in suspension cells

- **HYPE-5** dedicated to achieve High Yield Protein Expression in mammalian cells (CHO & HEK293 growing in suspension)
- **HYPE-293** dedicated to achieve High Yield Protein Expression in HEK293 cells growing in suspension

Purchaser Notification

Limited License

The purchase of the HYPE-CHO kit grants the purchaser a non-transferable, non-exclusive license to use the kit and/or its separate and included components (as listed in this protocol). This reagent is intended for in-house research only by the buyer. Such use is limited to the transfection of nucleic acids as described in the product manual. In addition, research only use means that this kit and all of its contents are excluded, without limitation, from resale, repackaging, or use for the making or selling of any commercial product or service without the written approval of OZ Biosciences. Separate licenses are available from OZ Biosciences for the express purpose of non-research use or applications of the HYPE-CHO kit. To inquire about such licenses, or to obtain authorization to transfer or use the enclosed material, contact us at OZ Biosciences. Buyers may end this License at any time by returning all HYPE-CHO kit reagents and documentation to OZ Biosciences, or by destroying all HYPE-CHO components. Purchasers are advised to contact OZ Biosciences with the notification that a HYPE-CHO kit is being returned in order to be reimbursed and/or to definitely terminate a license for internal research use only granted through the purchase of the kit(s). This document covers entirely the terms of the HYPE-CHO kit research only license, and does not grant any other express or implied license. The laws of the French Government shall govern the interpretation and enforcement of the terms of this License.

Product Use Limitations

HYPE-CHO kit and all of its components are developed, designed, intended, and sold for research use only. They are not to be used for human diagnostic or included/used in any drug intended for human use. All care and attention should be exercised in the use of the kit components by following proper research laboratory practices.

EUROPE & ASIA OZ Biosciences SAS

163 avenue de Luminy
Case 922, zone entreprise
13288 Marseille cedex 09
France

Ph: +33 (0) 486 948 516
Fax: +33 (0) 463 740 015

contact@ozbiosciences.com
order@ozbiosciences.com
tech@ozbiosciences.com



USA & CANADA OZ Biosciences INC

7975 Dunbrook Road
Suite B
San Diego CA 92126
USA

Ph: + 1-858-246-7840
Fax: + 1-855-631-0626

contactUSA@ozbiosciences.com
orderUSA@ozbiosciences.com
techUSA@ozbiosciences.com